# AMINO ACID SEQUENCE OF CYANOGEN BROMIDE FRAGMENT CB3 OF HOG PEPSIN 

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#### Abstract

On the basis of the knowledge of thermolytic, chymotryptic and substilisin peptides the amino acid sequence was determined of cyanogen bromide fragment CB3 representing the region between methionine residues I and II of pepsin: Thr-Gly-Ile-Leu-Gly-Tyr-Asp-Thr-Val-Gln-Val-Gly-Gly--Ile-Ser-Asp-Thr-Ast-Gln-Ile-Phc-Gly-Leu-Ser-Glu-Thr-Glu-Pro-Gly-Ser-Phe-Leu-Tyr-Tyr-Ala--Pro-Phe-Asp-Gly-Ile-Leu-Gly-Leu-Ala-Tyr-Pro-Ser-Ile-Scr-Ala-Ser-Gly-Ala-Thr-Pro-Val-Phe--Asp-Asn-Leu-Trp-Asp-Gln-Gly-Leu-Val-Ser-Gln-Asp-Leu-Phe-Scr-Val-Tyr-Leu-Ser-Ser-Asn--Asp-Asp-Ser-Gly-Ser-Val-Val-Leu-Leu-Gly-Gly-Ile-Asp-Ser-Ser-Tyr-Tyr-Thr-Gly-Ser-Leu-Asn--Trp-Val-Pro-Val-Ser-Val-Glu-Gly-Tyr-Trp-Gln-Ile-Thr-Leu-Asp-Ser-Jle-Thr-Mct.


In sequential studies on hog pepsin, carried out in this laboratory, fragments CB1 to CB6 were prepared by cyanogen bromide cleavage of S-sulfo-pepsin ${ }^{1}$. Tryptic digestion of aminoethylated pepsin${ }^{2}$ was employed as an alternative fragmentation procedure. The sequential analysis of the cyanogen bromide fragments ${ }^{3-5}$ provided the basic information for the determination of the complete and by now published ${ }^{6}$ amino acid sequence of the enzyme. As a result of incomplete cleavage at the carboxyl side of the first methionine residue (bond .. Met-Thr..) a longer fragment CB2, corresponding to the region of pepsin extending from the N -terminus to the methionine residue II and comprising segments corresponding to fragments CB4 and CB3 (ref. ${ }^{1}$ ), was isolated in addition to the products expected. For these reasons we were able to utilize for the investigation of the region between methionine residue I and II of pepsin both fragment CB3 and CB2, as well as tryptic fragment RAEP-tA 22 of aminoethylated pepsin ${ }^{2}$, also comprising this region.

This paper provides experimental data on the determination of the amino acid sequence of the remaining cyanogen bromide fragment CB3 representing the region of the polypeptide chain between methionine residues I and II.

## EXPERIMENTAL

## Material

Cyanogen bromide fragments CB2 and CB3 were prepared in earlier work ${ }^{1,5}$. The chemicals and materials used are described in earlier papers cited below.

Table I
Amino Acid Composition of Thermolytic Peptides Derived from the Region of Fragment CB3.
The peptides were analyzed after 20 -h hydrolysis; the values are not corrected. None of the peptides contained lysine, histidine, arginine, or half-cystine.

| Designation of peptide | Number of amino acid residues |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Asp | Thr | Ser | Glu | Pro | Gly | Ala | Val | Ile | Leu | Tyr | Phe | Hse ${ }^{\text {a }}$ | Trp |
|  |  |  | - |  |  |  |  |  |  |  |  |  |  |  |
| Thi | - | 2.0 | $1 \cdot 2$ | - | - | 3.0 | - | - | - | - | - | - | 0.8 | - |
| Th2 | - | 0.9 | - | - | - | 1.0 | - | - | - | - | - | - | $0 \cdot 7^{\text {b }}$ | - |
| Th3 | 10 | 0.9 | - | - | - | 1.0 | - | - | 1.0 | 1.0 | 0.8 | -.. | .- | - |
| Th4 | - | - | - | - | - | 1.0 | - | - | 0.8 | $1 \cdot 1$ | - | - | - | - |
| Th5 | $1 \cdot 0$ | 1.0 | - | - | - | - | - | - | - | - | 0.8 | - | - | - |
| Th6 | - | - | - | 1.0 | - | - | - | 1.0 | - | - | - | - | - | --- |
| Th7 | - | - | - | - | - | 1.9 | - | $1 \cdot 0$ | - | -- | - | - | - | - |
| Th8 | 2.0 | 0.9 | 1.0 | $1 \cdot 0$ | - | - | - | - | 1.0 | - | - | - | $\cdots$ | - |
| Th9 | - | - | - | - | - | $1 \cdot 0$ | -- | --- | 0.9 | - | - | $1 \cdot 1$ | - | - |
| Thl0 | - | 1.0 | 1.9 | $2 \cdot 0$ | $1 \cdot 0$ | $1 \cdot 1$ | - | - | - | 1.0 | - | 1.0 | -. | - |
| Thll | - | 1-1 | 2.0 | 1.9 | $1 \cdot 1$ | $1 \cdot 2$ | - | - | - | 0.7 | - | - | - | - |
| Th12 | -- | - | - | - | - | - | - | - | - | 1.0 | 1.0 | - | - | - |
| Th13 | - | - | - | - | 1.0 | - | 1.0 | - | - | - | 0.7 | - | - | - |
| Th14 | - | $\cdots$ | - | -- | 1.0 | - | 1.0 | - | - | - | - | - | - | - |
| Th15 | $1 \cdot 0$ | - | -- | -- | - | $1 \cdot 1$ | - | - | - | - | - | 1.0 | - | - |
| Th16 | - | - | - | - | - | 1.0 | - | - | 0.8 | $1 \cdot 1$ | - | - | - | - |
| Th17 | $1 \cdot 1$ | - | - | - | - | 1.9 | - | $\ldots$ | 0.8 | 1.0 | - | $1 \cdot 1$ | - | - |
| Th18 | - | -- | 0.9 | - | $1 \cdot 0$ | -- | 1.0 | - | - | 1.0 | 0.9 | -- | - | - |
| Th19 | - | - | 1.0 | - | 1.0 | - | $1 \cdot 0$ | - | - | - | 0.9 | - | - | - |
| Th20 | - | 1.3 | 1.8 | - | 1.0 | 1.1 | 2.0 | - | 0.9 | - | - | - | - | - |
| Th21 | $2 \cdot 1$ | 1.0 | 1.8 | - | 0.8 | 1.0 | 1.8 | 0.9 | 0.9 | - | - | 0.9 | - | - |
| Th22 | -- | - | $1 \cdot 8$ | - | - | $1 \cdot 3$ | 1.0 | - | 0.9 | - | - | - | - | -. |
| Th23 | $2 \cdot 1$ | 1.0 | - | - | 1.2 | - | 0.8 | 1.0 | - | - | - | 0.9 | - | - |
| Th24 | $2 \cdot 2$ | - | - | - | - | - | - | $1 \cdot 0$ | - | - | - | 1.0 | - | - |
| Th25 | $1 \cdot 1$ | - | - | 1.0 | .-- | 11 | - | - | - | 1.0 | - | -. | .... | $+^{\text {c }}$ |
| Th26 | $1 \cdot 2$ | - | 1.0 | $1 \cdot 0$ | - | - | - | $1 \cdot 1$ | - | 1.8 | - | - | - | - |
| Th27 | $1 \cdot 1$ | - | 1.0 | $1 \cdot 1$ | - | - | - | $1 \cdot 0$ | - | 0.9 | - | -- | -- | -. |
| Th28 | 1.0 | - | 0.9 | $1 \cdot 0$ | - | - | - | 08 | - | - | - | - | - | - |
| Th29 | - | - | 1.0 | - | - | - | - | - | - | - | - | 1.0 | - | - |
| Th30 | - | - | - | - | - | - | - | 1.0 | - | - | 0.9 | - | - | - |
| Th3I | $3 \cdot 3$ | - | $3 \cdot 8$ | - | - | $1 \cdot 2$ | - | 1.7 | - | 2.0 | - | - | - | - |
| Th32 | - | - | - | - | - | 2.0 | - | - | - | 1.0 | - | - | - | -- |
| Th33 | 1.3 | - | 1.9 | - | - | $2 \cdot 1$ | - | - | 0.9 | 0.8 | 1.1 | - | - | - |
| Th34 | $1 \cdot 1$ | - | 1.9 | - | - | - | - | - | 1.0 | - | 0.8 | - | - | - |
| Th35 | - | 1.0 | 1.0 | - | - | 1.0 | - | - | - | - | 0.8 | - | - | - |
| Th36 | $1 \cdot 0$ | - | - | - | $1 \cdot 1$ | - | - | 1.0 | - | 0.9 | - | - | - | $+^{c}$ |
| Th37 | - | - | 0.8 | - | - | - | - | 1.0 | - | - | - | - | - | - |
| Th38 | - | - | 0.9 | 1.1 | - | 1.0 | - | 2.0 | - | - | - | - | - | - |

Table I
(Continued)

| Designation of paptide | Number of amino acid residues |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Asp | Thr | Ser | Glu | Pro | Gly | Ala | Val | Ile | Leu | Tyr | Phe | Hse ${ }^{\text {a }}$ | Trp |
| Th39 | - | - | - | $1 \cdot 9$ | - | $1 \cdot 1$ | - | $1 \cdot 0$ | -- | - | 0.7 | - | - | $+^{c}$ |
| Th40 | - | - | - | 1.2 | - | $1 \cdot 1$ | - | 1.0 | - | - | 0.6 | - | - | - |
| Th41 | - | 1.2 | - | - | - | - | - | - | 1.0 | - | - | - | - | - |
| Th42 | $1 \cdot 1$ | - | $1 \cdot 0$ | - | - | - | - | - | - | 0.9 | - | - | - | - |
| Th43 | $1 \cdot 2$ | 1.9 | - | $1 \cdot 1$ | - | $1 \cdot 1$ | - | - | 1.0 | - | - | - | $0 \cdot 8^{\text {b }}$ | - |

${ }^{a}$ Hse denotes homoserine; ${ }^{b}$ methionine was determined in the peptide; ${ }^{c}$ the tryptophan content was determined qualitatively only.

## Methods

For chymotryptic digestion lyophilized fragment CB3 ( 350 mg ) was dissolved in 30 ml of water made alkaline with $0 \cdot 1 \mathrm{~m}-\mathrm{NH}_{4} \mathrm{HCO}_{3}$ (red color of phenol red added to the solution as an indicator). The substrate was cleaved by two portions of chymotrypsin (final weight ratio of enzyme to substrate $1: 50$ ) 4 h at $37^{\circ} \mathrm{C}$. The digest was fractionated by gel filtration on a column ( $65 \times$ $\times 5 \mathrm{~cm}$ ) of Sephadex G-15 equilibrated with $0.05 \mathrm{~m}-\mathrm{NH}_{4} \mathrm{HCO}_{3}$. Fractions ( $20 \mathrm{ml} / 15 \mathrm{~min}$ ) were evaluated by paper chromatography of aliquots $(0.2 \mathrm{ml})$ in the system 1-butanol-pyridine--acetic acid-water ( $15: 10: 3: 12, \mathrm{v} / \mathrm{v}$ ) (ref. ${ }^{7}$ ). Paper chromatography in this system and two electrophoretic procedures on Whatman No 3 or 3 MM paper were also used for the final purification of the peptides. The separation at pH 1.9 in the system formic acid-acetic acid-water ( $50: 150: 800, \mathrm{v} / \mathrm{v}$ ) at 4000 V was carried out in the horizontal arrangement in the apparatus constructed by Prusik and Štěpánek ${ }^{8}$. The separation at pH 5.6 in the system pyridine-acetic acid-water ( $5: 1: 494, \mathrm{v} / \mathrm{v}$ ) was performed at 1500 V in the vertical arrangement in the apparatus designed by Mikeš ${ }^{9}$.

The preparation and separation of the chymotryptic digest of fragment CB2 has been described in the preceding communication ${ }^{10}$, as well as the treatment of the chymotryptic digest of S -sulfo--pepsin ${ }^{11}$. The investigation of the thermolysin digest of fragment CB2 (ref. ${ }^{10}$ ) and of the thermolysin digest of S-sulfo-pepsin ${ }^{12}$ have also been described before. The amino acid compositions and sequences of those peptides from the digests which fall into the region of fragment CB3 are given in Tables I-IV. Peptides obtained in duplicate are listed once only, in order of their location in the sequence of fragment CB3. Subtilisin peptides, given in the scheme in Fig. 1, were isolated from the digest of tryptic fragment RAEP-tA. 22 of aminoethylated pepsin ${ }^{13}$.

Amino acid analyses were performed on 20 - and 70 -h hydrolysates (at $110^{\circ} \mathrm{C}$ ) by the method of Spackman and coworkers ${ }^{14}$ as modified by Benson and Patterson ${ }^{15}$ in Model 6020 Amino Acid Analyzer, manufactured by the Instrument Development Workshop, Czechoslovak Academy of Sciences, Prague. Homoserine lactone was converted ${ }^{16}$ to homoserine before the analysis. The amino acid sequences of peptides were determined by Edman degradation ${ }^{17}$; the fenylthiohydantoins were analyzed by thin-layer chromatography on silica gel. In certain cases the chymo-

## Table II

Sequence of Thermolytic Peptides Derived from the Region of Fragment CB3
Designation
of peptide
Amino acid sequence of peptide

| Th1 | Gly-Thr-Gly-Ser-Hse-Thr-Gly |
| :---: | :---: |
| Th2 | Met-Thr-Gly |
| Th3 | Ile-Leu-Gly-Tyr-Asp-Thr |
| Th4 | Ile-Leu-Gly |
| Th5 | Tyr-Asp-Thr |
| Th6 | Val-Gln |
| Th7 | Val-Gly-Gly |
| Th8 | Ile-Ser-Asp-Thr-Asn-Glo |
| Th9 | Ile-Phe-Gly |
| Th10 | Leu(Ser,Glx,Thr,Glx,Pro,Gly,Ser,Phe) |
| Th11 | Leu(Ser,Glx, Thr,Glx, Pro,Gly,Ser) |
| Th12 | Leu-Tyr |
| Th13 | Tyr-Ala-Pro |
| Th14 | Ala-Pro |
| Th15 | Phe(Asp, Giy) |
| Th16 | Ile-Leu-Gly |
| Th17 | (Phe, Asx, Gly, Ile, Leu, Gly) |
| Th18 | Leu-Ala-Tyr-Pro-Scr |
| Th19 | Ala-Tyr-Pro-Ser |
| Th20 | Ilc-Scr-Ala-Scr-Gly-Ala-Thr-Pro |
| Th21 | Ile(Ser,Ala,Ser,Gly)(Ala, Thr, Pro, Val, Phe,Asx, Asx) |
| Th22 | (Ile,Ser,Ala,Ser,Gly) |
| Th23 | (Ala,Thr,Pro, Val,Phe,Asx,Asx) |
| Th24 | (Val,Phe,Asx,Asx) |
| Th25 | Leu-Trp-Asp-Gln-Gly |
| Th26 | Leu-Val-Ser-Gln-Asp-Leu |
| Th27 | Leu(Val,Ser,Glx,Asx) |
| Th28 | Val(Ser,Glx,Asx) |
| Th29 | Phe-Ser |
| Th30 | Val-Tyr |
| Th31 | Leu-Ser-Ser-Asn-Asp-Asp(Ser,Gly,Ser,Val,Val,Leu) |
| Th32 | Leu-Gly-Gly |
| Th33 | Lcu(Gly,Gly,Ilc,Asx,Ser,Ser,Tyr) |
| Th34 | Ile(Asx,Ser,Ser, Tyr) |
| Th35 | Tyr-Thr-Gly-Ser |
| Th36 | Leu-Asn-Trp-Val-Pro |
| Th37 | Val-Ser |
| Th38 | Val(Ser,Val,Glu,Gly) |
| Th39 | $\mathrm{Val}(\mathrm{Glx}, \mathrm{Gly}$, Tyr, Trp)Gln |
| Th40 | Val-Glu(Gly, Tyr) |
| Th41 | Ile-Thr |
| Th42 | Leu-Asp-Ser |
| Th43 | Ile(Thr,Met,Asx,Gly,Glx,Thr) |

tryptic peptides were assigned C-terminal end groups with respect to the specificity of cleavage by this enzyme. Other details of the isolation and characterization of the peptides are described in the preceding papers ${ }^{11-13}$.

## RESULTS AND DISCUSSION

Fragment CB3 is formed by cyanogen bromide cleavage of S-sulfo-pepsin, at methionine residues I and II (ref. ${ }^{1}$ ). As a result of incomplete cleavage of the bond involving methionine I this part of the pepsin chain is contained also in the product cleaved

Table III
Amino Acid Composition of Chymotryptic Peptides Derived from the Region of Fragment CB3
The peptides were analyzed after 20 h hydrolysis; the values are not corrected. None of the peptides contained lysine, histidine, arginine, or half-cystine.

| Designation of peptide | Number of amino acid residues |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Asp | Thr | Ser | Glu | Pro | Gly | Ala | Val | Ile | Leu | Tyr | Phe | Hse | Trp |
| Cl | - | 1.9 | $1 \cdot 0$ | - | - | 3.6 | - | - | $1 \cdot 0$ | $1 \cdot 0$ | 0.5 | - | $1 \cdot 2$ | - |
| C2 | - | $2 \cdot 0$ | $1 \cdot 0$ | - | - | 2.9 | - | - | $1 \cdot 0$ | 1.0 | - | - | $1 \cdot 1$ | - |
| C3 | $3 \cdot 1$ | 2.0 | 1.0 | $2 \cdot 2$ | - | $2 \cdot 2$ | - | $2 \cdot 0$ | 1.7 | - | -- | 0.8 | - | - |
| C4 | - | $1 \cdot 1$ | 2.0 | 2.0 | $1 \cdot 1$ | 2.0 | - | - | - | $1 \cdot 1$ | - | 1.0 | - | - |
| C5 | - | 1.0 | $2 \cdot 1$ | 2.0 | 1.0 | 2.0 | - | - | - | 1.9 | 0.8 | 1.0 | - | - |
| C6 | - | 1.0 | 1.7 | 2.0 | 1.0 | 2.0 | - | - | - | $2 \cdot 1$ | - | 1.0 | - | - |
| C7 | - | - | - | - | - | - | - | - | $\cdots$ | 1.0 | 0.8 | - | - | - |
| C8 | -. | -. | - | - | $1 \cdot 1$ | - | 1.0 | - | - | - | 1.0 | 1.0 | - | - |
| C9 | $1 \cdot 1$ | - | --.. | - | $1 \cdot 1$ | $1 \cdot 1$ | 1.1 | $\cdots$ | 0.9 | 1.0 | 0.6 | 1.0 | - | - |
| C10 | - | - | - | -- | 1.0 | - | 1.0 | - | - | - | -- | 0.9 | - | - |
| C11 | 1.0 | - | - | - | - | $2 \cdot 1$ | - | - | 0.9 | 2.0 | - | - | - | $\cdots$ |
| C12 | 4.8 | 1.4 | 4.0 | 2.5 | $2 \cdot 1$ | 2.6 | 3.0 | $2 \cdot 2$ | 1.2 | 3.4 | 0.9 | $2 \cdot 1$ | - | + ${ }^{\text {a }}$ |
| C13 | 4.1 | 1.2 | 1.9 | $2 \cdot 3$ | 1.2 | 2.2 | 1.8 | 2.0 | - | 3.0 | - | 1.9 | - | + ${ }^{\text {a }}$ |
| C14 | $2 \cdot 1$ | - | $1 \cdot 3$ | 1.7 | - | $1 \cdot 1$ | - | $1 \cdot 1$ | - | 1.9 | - | $0 \cdot 8$ | - | - |
| C15 | $1 \cdot 1$ | - | 1.1 | 1.0 | -- | - | - | 0.9 | - | 0.9 | - | 0.8 | - | - |
| C16 | - | - | 0.9 | - | - | - | $\cdots$ | 1.0 | - | - | 0.9 | - | - | - |
| C17 | 3.0 | - | 3.7 | - | - | $1 \cdot 1$ | - | 1.5 | - | 2.0 | - | - | - | - |
| C18 | 1.3 | - | 2.0 | - | - | 2.0 | - | - | 1.0 | $1 \cdot 1$ | 1.6 | -- | - | - |
| C19 | $1 \cdot 1$ | 0.9 | 1.0 | - | - | 1.0 | - | - | - | $1 \cdot 0$ | - | - | - | + |
| C20 | - | - | 0.9 | 0.9 | 1.0 | 1.0 | - | 2.8 | - | - | 1.0 | - | - | + |
| C21 | $1 \cdot 0$ | 1.0 | $1 \cdot 0$ | - | - | - | - | - | 0.9 | - | - | - | 1.2 | - |

[^0]Table IV
Sequence of Chymotryptic Peptides Derived from the Region of Fragment CB3

```
Designation
    of peptide
```

Amino acid sequence of peptide

| Cl | Gly-Thr-Gly(Ser,Hse,Thr,Gly)(Ilc, Lcu,Gly)Tyr |
| :---: | :---: |
| C2 | Gly(Thr,Gly,Ser,Hsc, Thr,Gly,Ile)Leu |
| C3 | Asp-Thr-Val-Gln-Val-Gly-Gly-Ile-Ser-Asp(Thr,Asx,Glx,Ile)Phe |
| C4 | Gly-Leu-Ser-Glu-Thr-Glu-Pro-Gly-Ser-Phe |
| C5 | Gly(Leu,Ser,Glx,Thr,Glx,Pro,Gly,Scr)Phe-Leu-Tyr |
| C6 | (Gly,Leu,Ser,Glx, Thr, Glx, Pro,Gly,Ser,Phe,Leu) |
| C7 | Leu-Tyr |
| C8 | Tyr-Ala-Pro-Phe |
| C9 | (Tyr,Ala, Pro, Phe, Asx, Gly, Ilc, Leu) |
| C10 | (Ala, Pro)Phe |
| Cl1 | Asp-Gly-Ile-Leu-Gly-Lcu |
| C12 | Ala-Tyr-Pro-Ser-Ile-Srr-Ala(Ser ${ }_{2}, \mathrm{Gly}_{2}, \mathrm{Ala}, \mathrm{Thr}$, Pro, $\mathrm{Val}_{2}, \mathrm{Phc}_{2}$, Asx $_{4}$, Leu $_{3}, \mathrm{Trp}$, , Glx ${ }_{2}$ ) |
| C13 | Ala-Ser-Gly-Ala-Thr-Pro-Val-Phe-Asp(Asx,Leu,Trp,Asx,Glx,Gly,Leu,Val,Ser, ,Glx,Asx, Leu,Phe) |
| C14 | Asp-Gln-Gly-Leu-Val-Ser-Gln-Asp-Leu-Phe |
| C15 | (Val,Ser,Glx,Asx Leu)Phe |
| C16 | Ser-Val-Tyr |
| C 17 | Leu-Ser-Ser-Asn-Asp-Asp-Ser-Gly-Ser-Val-Val-Leu |
| C18 | (Leu, Gly, Gly, (le,Asx,Ser,Ser,Tyr)Tyr |
| C19 | Thr-Gly-Ser-Leu-Asn-Trp |
| C20 | (Val,Pro,Val,Ser, Val,Glu,Gly, Tyr)Trp |
| C21 | Asp-Scr-Ile-Thr-Hse |

Table V
Amino Acid Composition of Pragmert CB3
The values are given as number of amino acid residues in the molecule of the fragment. Arginine, lysine, histidine, a hall-cystine were not found in the fragment.

Fragment Asp Thr Ser Glu Pro Gly Ala Val Ile Leu Tyr Phe Hse Trp

| CB3 (rcf. ${ }^{5}$ ) | 14.0 | $7 \cdot 8^{\text {a }}$ | $16.9{ }^{\text {a }}$ | 8.2 | 4.6 | 14.6 | $4 \cdot 1$ | $9 \cdot 8{ }^{\text {b }}$ | $7.9{ }^{\text {b }}$ | 12.7 | $7 \cdot 4$ | $5 \cdot 0$ | $0 \cdot 8$ | $+^{c}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| In sequence of CB3 | 14 | 8 | 17 | 8 | 5 | 15 | 4 | 10 | 8 | 13 | 8 | 5 | 1 | 3 |

[^1]incompletely (fragment CB2) in which the N -terminal fragment CB4 is linked to fragment CB3 following in the chain. Fragment CB3 was obtained during the basic fractionation of the cyanogen bromide digest by gel filtration; this preparation containing contaminants was characterized ${ }^{1}$ by the $N$-terminal sequence Thr-Gly-Ile--Leu-Gly-Tyr-... and by the C-terminal sequence $\operatorname{Ser}(\mathrm{lle}, \mathrm{Thr})$ Hse. The carboxyl terminus of fragment CB3 is identical with the C-terminus of fragment CB2, determined as Ile-Thr-Hse. The isolation of the corresponding homoserine peptide from the chymotryptic digest of fragment CB2 (ref. ${ }^{10}$ ) permitted the carboxyl terminal region of fragment CB3 and CB2 to be formulated as Asp-Ser-Ile-Thr-Hse (ref. ${ }^{1}$ ). Fragment CB3 was purified by ion cxchange chromatography ${ }^{5}$; the amino acid composition of this preparation is given in Table V. The tryptophan content (Table V) was not determined directly, it is known, however, from the analysis of fragment RAEP-tA 22 ( 2.9 residues, ref. ${ }^{2}$ ), whose tcrminal parts exceeding fragment CB3 do not contain tryptophan ${ }^{5}$. The N -terminal sequence of the fragment, Thr-Gly-lle--Leu-Gly-Tyr-..., was verified by manual Edman degradation of pure fragment CB3. Because of the absence of lysine, argininc. and half-cystine (convertible into the aminoethylcysteinc residue, sensitive to trypsin) in the part of the pepsin chain overlapping fragment CB3 this part of the chain was not cleaved by tryptic digestion of aminoethylated pepsin and a large fragment RAEP-tA 22 (ref. ${ }^{2}$ ) derived from this site was isolated. For the same reason tryptic hydrolysis could not be employed for sequential investigation of fragment CB3.

The amino acid sequence of the fragment (Fig. 1) was derived mainly from information afforded by chymotryptic, thermolytic, and subtilisin peptides. Data on the chymotryptic peptides obtained by the digestion of fragment CB3 were complemented by the alignment of peptides from the chymotryptic digest ${ }^{10}$ of fragment CB2 (after elimination of peptides incorporated into the sequence ${ }^{5}$ of fragment CB4) and, knowing the structures of the remaining cyanogen bromide fragments ${ }^{3-5}$, also by selected peptides from the chymotryptic hydrolysate of S-sulfo-pepsin ${ }^{11}$. Likewise, after the determination of the sequence of fragment $C B 4$, the peptides remaining from the thermolytic digest ${ }^{10}$ of fragment CB 2 could be unambiguously incorporated into the region of CB3 and it became possible to also align selected peptides from the thermolysin digest of S-sulfo-pepsin ${ }^{12}$. Tryptic fragment RAEP-tA 22 (ref. ${ }^{2}$ ) served as a source for the preparation of subtilisin peptides. The subtilisin digest is described in more detail in the adjoining paper ${ }^{13}$; only selected peptides from this digest, complementing data obtained with other digests, are shown in the scheme in Fig. 1. In a number of cases the sequential characterization of the peprides (Table II and IV) was omitted wherever the information had been obtained with other peptides.

Unambiguous sequential overlaps (Fig. 1) permitted the amino acid sequence of the fragment to be arranged into four sections:

|  |  |
| :---: | :---: |
|  |  |
|  |  |


Asn-Gln-Ile-Phe-Gly-Ieu-Ser-Glu-Thr-Glu-Pro-Gly-Ser-Phe-Leu-Tyr-Iyr-Ala-Pro-Phe-Asp-Gly-

 to the adjoining paper ${ }^{13}$. The Edman degradation steps are marked $\rightarrow$, the results of carboxypeptidase A cleavage $\leftarrow$.






|  |  |  |  |  |  |  |  |  |  |
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| I | Thr-Gly-Ile | Ser-Val-Tyr | (res. 1-74) |
| :---: | :---: | :---: | :---: |
| II | Leu-Ser-Ser | Val-Val-Leu | (res. 75-86) |
| III | Leu-Gly-Gly | Gln-Ile-Thr | (res. 87-113) |
| IV | Leu-Asp-Ser- |  | (res. 114-119) |

We were not able to join together the above regions by overlapping peptides. The total sum of amino acid residues in these noneverlapping sequential regions equals the amino acid composition of fragment CB3 (Table V). The peptides from the individual enzymatic digests comply with this arrangement of sequential regions and do not suggest the existence of overlaps or, by contrast, a loss of peptides where these regions are linked to one another (residues $74 / 75,86 / 87$, and 113/114). The N -terminal sequence of region I is identical to the N -terminal sequence of fragment CB3; sequence IV, containing the homoserine residue, represents the carboxyl terminus of the fragment. The order of the middle regions II and III was established from the results of carboxypeptidase A degradation of fragments CB3 and CB2 (ref. ${ }^{1}$ ). During prolonged cleavage the quantity of threonine and isoleucine liberated exceeds the quantity of C-terminal homoserine; it may thus be assumed that the short sequential region IV (Leu-Asp-Ser-Ile-Thr-Hse) is preceded by a threonine and isoleucine residue. This assumption is in accordance with the carboxyl terminus of region III, ..GIn-Ile-Thr, since region II is C-terminated by ..Val-Val-Leu. As evidence supporting the location of region II after the N -terminal sequence I may be adducted the fact that the bond at the amino terminus of leucine No 75 is susceptible to cleavage by both chymotrypsin and bromosuccinimide ${ }^{18}$. This observation requires the presence of a tyrosine residue at the amino side of leucine 155 since the complete knowledge of the sequence around the three tryptophans of fragment CB 3 excludes this residue as an alternative. Likewise, the sensitivity of the bond between residues 86 and 87 to chymotrypsin is compatible with the proposed bond Leu(86)-Leu(87).

The sequence of fragment CB2A, determined by Sepulveda and coworkers ${ }^{19}$ is in complete agreement with our sequence of fragment CB3 (Fig. 1) which has already been reported in the paper describing the complete amino acid sequence of hog pepsin${ }^{6}$; partial data pertaining to this part of the molecule were also published by Ostoslavskaya and coworkers ${ }^{20}$. Likewise the results reported by Revina and coworkers ${ }^{21}$, accounting for residues $100-119$ of this fragment, confirm our sequence.

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[^0]:    ${ }^{a}$ The tryptophan content was determined qualitatively only.

[^1]:    a Values extrapolated to zero time of hydrolysis; ${ }^{b}$ values determined after 70-h hydrolysis;
    ${ }^{c}$ the tryptophan content was determined qualitatively only.

