# AMINO ACID SEQUENCE OF CYANOGEN BROMIDE FRAGMENT CB3 OF HOG PEPSIN

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On the basis of the knowledge of thermolytic, chymotryptic and substilisin peptides the amino acid sequence was determined of cyanogen bromide fragment CB3 representing the region between methionine residues I and II of pepsin: Thr-Gly-Ile-Leu-Gly-Tyr-Asp-Thr-Val-Gln-Val-Gly-Gly-Ile-Ser-Asp-Thr-Asn-Gln-Ile-Phe-Gly-Leu-Ser-Glu-Thr-Glu-Pro-Gly-Ser-Phe-Leu-Tyr-Tyr-Ala-Pro-Phe-Asp-Gly-Ile-Leu-Gly-Leu-Val-Ser-Gln-Asp-Leu-Ple-Ser-Val-Tyr-Leu-Ser-Asn-Asp-Asn-Leu-Tyr-Asp-Gln-Gly-Leu-Val-Ser-Gln-Asp-Leu-Ple-Ser-Val-Tyr-Leu-Ser-Ser-Asn-Asp-Asp-Ser-Gly-Ser-Val-Val-Leu-Leu-Gly-Gly-Ile-Asp-Ser-Ser-Tyr-Tyr-Thr-Gly-Ser-Leu-Asp--Trp-Val-Ser-Val-Glu-Gly-Tyr-Trp-Gln-Ile-Thr-Leu-Asp-Ser-Ile-Thr-Met.

In sequential studies on hog pepsin, carried out in this laboratory, fragments CB1 to CB6 were prepared by cyanogen bromide cleavage of S-sulfo-pepsin<sup>1</sup>. Tryptic digestion of aminoethylated pepsin<sup>2</sup> was employed as an alternative fragmentation procedure. The sequential analysis of the cyanogen bromide fragments<sup>3-5</sup> provided the basic information for the determination of the complete and by now published<sup>6</sup> amino acid sequence of the enzyme. As a result of incomplete cleavage at the carboxyl side of the first methionine residue (bond ...Met-Thr...) a longer fragment CB2, corresponding to the region of pepsin extending from the N-terminus to the methionine residue II and comprising segments corresponding to fragments CB4 and CB3 (ref.<sup>1</sup>), was isolated in addition to the products expected. For these reasons we were able to utilize for the investigation of the region between methionine residue I and II of pepsin both fragment CB3 and CB2, as well as tryptic fragment RAEP-tA 22 of aminoethylated pepsin<sup>2</sup>, also comprising this region.

This paper provides experimental data on the determination of the amino acid sequence of the remaining cyanogen bromide fragment CB3 representing the region of the polypeptide chain between methionine residues I and II.

# EXPERIMENTAL

#### Material

Cyanogen bromide fragments CB2 and CB3 were prepared in earlier work<sup>1,5</sup>. The chemicals and materials used are described in earlier papers cited below.

# TABLE I

Amino Acid Composition of Thermolytic Peptides Derived from the Region of Fragment CB3. The peptides were analyzed after 20-h hydrolysis; the values are not corrected. None of the peptides contained lysine, histidine, arginine, or half-cystine.

Designation					Nu	mber	of ami	no ac	id resi	idues				
of peptide	Asp	Thr	Ser	Glu	Pro	Gly	Ala	Val	Ile	Leu	Tyr	Phe	Hse <sup>a</sup>	Trp
		2.0			_	3.0			nderson <b>L</b> abert					
Thi Th2	_	2·0 0·9	1.2	_		3.0		_				_	0·8 0·7 <sup>b</sup>	
Th2 Th3	1.0	0.9	_	_		1.0			1.0	1.0	0.8			
Th4	1.0	0.9			_	1.0		_	0.8	1.1	0.9			_
Th5	1.0	1.0	_	_	_		_	_			0.8	_		_
Th6		_		1.0	_		_	1.0			_	_		
Th7		_			_	1.9		1.0		_	_			
Th8	2.0	0.9	· 1·0	1.0	_			_	1.0	_		_		
Th9	2.0	0.9	1.0		_	1.0			0.9	_		1.1		_
Th10		1.0	1.9	2.0	1.0	1.1	_	_		1.0	_	1.0		
Thii	_	1.1	2.0	1.9	1.1	1.2	_		_	0.7	_			
Th12	_	_	-		_		_		_	1.0	1.0	_		_
Th12	_		_	_	1.0	_	1.0				0.7	_		
Th14	_		_		1.0	-	1.0	_	_	_	_			_
Th15	1.0	_			_	1.1			_			1.0		
Th16			_	_		1.0	_		0.8	1.1	_		_	
Th17	1.1					1.9	·		0.8	1.0	_	1.1	_	
Th18			0.9	_	1.0		1.0	_	_	1.0	0.9			_
Th19		_	1.0		1.0	_	1.0			_	0.9	_	_	
Th20		1.3	1.8	_	1.0	1.1	2.0		0.9		_	_	_	
Th21	2.1	1.0	1.8		0.8	1.0	1.8	0.9	0.9	_	_	0.9		_
Th22		_	1.8		_	1.3	1.0	_	0.9	_				
Th23	2.1	1.0		_	1.2	_	0.8	1.0		_		0.9		
Th24	2.2	-			_			1.0				1.0		
Th25	1.1		·	1.0		1.1	_	_	_	1.0	_			+. °
Th26	1.2		1.0	1.0		_		1-1		1.8		_	_	
Th27	1.1	_	1.0	1.1	_	_		1.0	_	0.9				
Th28	1.0		0.9	1.0				0.8		_		_	_	_
Th29	_	_	1.0	—		_		_	_		_	1.0		_
Th30				-			_	1.0	—	-	0.9			
Th31	3.3		3.8		-	1.2		1.7	_	2.0		_		
Th32						2.0		_		1.0			-	
Th33	1.3		1.9	_	-	2.1	_	_	0.9	0.8	1.1			
Th34	1.1		1.9			_	_	_	1.0		0.8	_	_	
Th35		1.0	1.0			1.0	-	_		_	0.8	_		_
Th36	1.0	_			1.1		_	1.0	_	0.9	-			+ °
Th37			0.8	—	_	-	_	1.0	_			-		-
Th38	-	_	0.9	1.1		1.0	1.00	2.0		_		_	_	-

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## Amino Acid Sequence of Cyanogen Bromide

TABLE	I
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### (Continued)

Designation of peptide		Number of amino acid residues														
	Asp	Thr	Ser	Glu	Pro	Gly	Ala	Val	Ile	Leu	Tyr	Phe	Hseª	Trp		
Th39	_	_	_	1.9	_	1.1		1.0		_	0.7	_		·+ •		
Th40	_	_		1.2		1.1	_	1.0	_		0.6		-			
Th41	_	1.2	_	_		_	_		1.0	_	_	_				
Th42	1.1		1.0		<u> </u>	_		-	_	0.9				_		
Th43	1.2	1.9		1.1	_	1.1	_	_	1.0	_	_	_	0·8 <sup>b</sup>			

<sup>a</sup> Hse denotes homoserine; <sup>b</sup> methionine was determined in the peptide; <sup>c</sup> the tryptophan content was determined qualitatively only.

#### Methods

For chymotryptic digestion lyophilized fragment CB3 (350 mg) was dissolved in 30 ml of water made alkaline with 0.1M-NH<sub>4</sub>HCO<sub>3</sub> (red color of phenol red added to the solution as an indicator). The substrate was cleaved by two portions of chymotrypsin (final weight ratio of enzyme to substrate 1: 50) 4 h at 37°C. The digest was fractionated by gel filtration on a column (65 × × 5 cm) of Sephadex G-15 equilibrated with 0.05M-NH<sub>4</sub>HCO<sub>3</sub>. Fractions (20 ml/15 min) were evaluated by paper chromatography of aliquots (0·2 ml) in the system 1-butanol-pyridine--acetic acid-water (15:10:3:12, v/v) (ref.<sup>7</sup>). Paper chromatography in this system and two electrophoretic procedures on Whatman NO 3 or 3 MM paper were also used for the final purification of the peptides. The separation at pH 1·9 in the system formic acid-acetic acid-water (50:150:800, v/v) at 4000 V was carried out in the horizontal arrangement in the apparatus constructed by Prusik and Štěpánek<sup>8</sup>. The separation at pH 5·6 in the system pyridine-acetic acid-water (5:1:494, v/v) was performed at 1500 V in the vertical arrangement in the apparatus designed by Mikes<sup>9</sup>.

The preparation and separation of the chymotryptic digest of fragment CB2 has been described in the preceding communication<sup>10</sup>, as well as the treatment of the chymotryptic digest of S-sulfopepsin<sup>11</sup>. The investigation of the thermolysin digest of fragment CB2 (ref.<sup>10</sup>) and of the thermolysin digest of S-sulfo-pepsin<sup>12</sup> have also been described before. The amino acid compositions and sequences of those peptides from the digests which fall into the region of fragment CB3 are given in Tables I—IV. Peptides obtained in duplicate are listed once only, in order of their location in the sequence of fragment CB3. Subtilisin peptides, given in the scheme in Fig. 1, were isolated from the digest of tryptic fragment RAEP-LA 22 of aminoethylated pepsin<sup>13</sup>.

Amino acid analyses were performed on 20- and 70-h hydrolysates (at 110°C) by the method of Spackman and coworkers<sup>14</sup> as modified by Benson and Patterson<sup>15</sup> in Model 6020 Amino Acid Analyzer, manufactured by the Instrument Development Workshop, Czechoslovak Academy of Sciences, Prague. Homoserine lactone was converted<sup>16</sup> to homoserine before the analysis. The amino acid sequences of peptides were determined by Edman degradation<sup>17</sup>; the fenylthio-hydantoins were analyzed by thin-layer chromatography on silica gel. In certain cases the chymo-

# TABLE II

Sequence of Thermolytic Peptides Derived from the Region of Fragment CB3

Designation of peptide	Amino acid sequence of peptide						
Th1	Gly-Thr-Gly-Ser-Hse-Thr-Gly						
Th2	Met-Thr-Gly						
Th3	Ilc-Leu-Gly-Tyr-Asp-Thr						
Th4	Ile-Leu-Gly						
Th5	Tyr-Asp-Thr						
Th6	Val-Gln						
Th7	Val-Gly-Gly						
Th8	Ile-Ser-Asp-Thr-Asn-Gln						
Th9	Ile-Phe-Gly						
Th10	Lcu(Ser,Glx,Thr,Glx,Pro,Gly,Ser,Phe)						
Th11	Leu(Ser,Glx,Thr,Glx,Pro,Gly,Ser)						
Th12	Leu-Tyr						
Th13	Tyr-Ala-Pro						
Th14	Ala-Pro						
Th15	Phe(Asp, Gly)						
Th16	Ile-Leu-Gly						
Th17	(Phe, Asx, Gly, Ile, Leu, Gly)						
Th18	Leu-Ala-Tyr-Pro-Ser						
Th19	Ala-Tyr-Pro-Ser						
Th20	Ilc-Scr-Ala-Scr-Gly-Ala-Thr-Pro						
Th21	Ile(Ser,Ala,Ser,Gly)(Ala,Thr,Pro,Val,Phe,Asx,Asx)						
Th22	(Ile,Ser,Ala,Ser,Gly)						
Th23	(Ala,Thr,Pro,Val,Phe,Asx,Asx)						
Th24	(Val,Phe,Asx,Asx)						
Th25	Leu-Trp-Asp-Gin-Gly						
Th26	Leu-Val-Ser-Gln-Asp-Leu						
Th27	Leu(Val,Ser,Glx,Asx)						
Th28	Val(Ser,Glx,Asx)						
Th29	Phe-Ser						
Th30	' Val-Tyr						
Th31	Leu-Ser-Ser-Asn-Asp-Asp(Ser,Gly,Ser,Val,Val,Leu)						
Th32	Leu-Gly-Gly						
Th33	Lcu(Gly,Gly,Ilc,Asx,Ser,Ser,Tyr)						
Th34	Ile(Asx,Ser,Ser,Tyr)						
Th35	Tyr-Thr-Gly-Ser						
Th36	Leu-Asn-Trp-Val-Pro						
Th37	Val-Ser						
Th38	Val(Ser, Val, Glu, Gly)						
Th39	Val(Glx,Gly,Tyr,Trp)Gln						
Th40	Val-Glu(Gly,Tyr)						
Th41	Ile-Thr						
Th42	Leu-Asp-Ser						
Th43	Ile(Thr, Met, Asx, Gly, Glx, Thr)						

tryptic peptides were assigned C-terminal end groups with respect to the specificity of cleavage by this enzyme. Other details of the isolation and characterization of the peptides are described in the preceding papers<sup>11-13</sup>.

# RESULTS AND DISCUSSION

Fragment CB3 is formed by cyanogen bromide cleavage of S-sulfo-pepsin, at methionine residues I and II (ref.<sup>1</sup>). As a result of incomplete cleavage of the bond involving methionine I this part of the pepsin chain is contained also in the product cleaved

## TABLE III

Amino Acid Composition of Chymotryptic Peptides Derived from the Region of Fragment CB3 The peptides were analyzed after 20 h hydrolysis; the values are not corrected. None of the peptides contained lysine, histidine, arginine, or half-cystine.

Designation	Number of amino acid residues													
of peptide	Asp	Thr	Ser	Glu	Pro	Gly	Ala	Val	Ile	Leu	Tyr	Phe	Hse	Tr
CI	_	1.9	1.0		_	3.6	_	_	1.0	1.0	0.5		1.2	
C2		2.0	1.0			2.9			1.0	1.0	_	_	1.1	_
C3	3.1	2.0	1.0	2.2		2.2	_	2.0	1.7			0.8	_	_
C4	_	1.1	2.0	2.0	1.1	2.0			_	1.1	_	1.0		
C5		1.0	2.1	2.0	1.0	2.0		_		1.9	0.8	1.0	_	-
C6	_	1.0	1.7	2.0	1.0	2.0			_	2.1	-	1.0		~
C7	_	_	_	_	_	A17.78			1.000	1.0	0.8			-
C8			_		1.1	-	1.0	_			1.0	1.0		_
C9	1.1	_		_	1.1	1.1	I · 1		0.9	1.0	0.6	1.0		-
C10		_	_	-	1.0		1.0		-	-		0.9		-
C11	1.0		_	_		2.1		_	0.9	2.0	_		_	-
C12	4.8	1.4	4·0	2.5	2.1	2.6	3.0	2.2	1.2	3.4	0.9	2.1		~
C13	4.1	1.2	1.9	2.3	1.2	2.2	1.8	2.0	_	3.0		1.9	wood	-
C14	2.1	_	1.3	1.7	_	1.1		1.1	_	1.9		0.8		_
C15	1.1	_	1.1	1.0	1011-104	_		0.9		0.9		0.8		-
C16	_		0.9	_		amand.	a	1.0			0.9	******	_	-
C17	3.0		3.7		_	1.1	_	1.5		2.0	_	_		-
C18	1.3	_	2.0			2.0		_	1.0	1.1	1.6			-
C19	1.1	0.9	1.0			1.0	·	_		1.0		_	_	-
C20			0.9	0.9	1.0	1.0	_	2.8		_	1.0	_		-
C21	1.0	1.0	1.0		_		_	_	0.9		_		1.2	-

<sup>a</sup> The tryptophan content was determined qualitatively only.

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# TABLE IV

Sequence of Chymotryptic Peptides Derived from the Region of Fragment CB3

Designation of peptide	Amino acid sequence of peptide
CI	Gly-Thr-Gly(Ser,Hse,Thr,Gly)(Ile,Leu,Gly)Tyr
C2	Gly(Thr,Gly,Ser,Hsc,Thr,Gly,Ile)Leu
C3	Asp-Thr-Val-Gln-Val-Gly-Gly-Ile-Ser-Asp(Thr,Asx,Glx,Ile)Phe
C4	Gly-Leu-Ser-Ġlu-Thr-Glu-Pro-Gly-Ser-Phe
C5	Gly(Leu,Ser,Glx,Thr,Glx,Pro,Gly,Scr)Phc-Lcu-Tyr
C6	(Gly,Leu,Ser,Glx,Thr,Glx,Pro,Gly,Ser,Phe,Leu)
C7	Leu-Tyr
C8	Tyr-Ala-Pro-Phe
C9	(Tyr,Ala,Pro,Phe,Asx,Gly,Ilc,Leu)
C10	(Ala,Pro)Phe
CH	Asp-Gly-Ile-Leu-Gly-Leu
C12	Ala-Tyr-Pro-Ser-Ile-Ser-Ala(Ser <sub>2</sub> ,Gly <sub>2</sub> ,Ala,Thr,Pro,Val <sub>2</sub> ,Phc <sub>2</sub> ,Asx <sub>4</sub> ,Leu <sub>3</sub> ,Trp, ,Glx <sub>2</sub> )
C13	Ala-Ser-Gly-Ala-Thr-Pro-Val-Phe-Asp(Asx,Leu,Trp,Asx,Glx,Gly,Leu,Val,Ser, ,Glx,Asx,Leu,Phe)
C14	Asp-Gln-Gly-Leu-Val-Ser-Gln-Asp-Leu-Phe
C15	(Val,Ser,Glx,Asx Leu)Phe
C16	Ser-Val-Tyr
C17	Leu-Ser-Ser-Asn-Asp-Asp-Ser-Gly-Ser-Val-Val-Leu
C18	(Leu, Gly, Gly, Ile, Asx, Ser, Ser, Tyr)Tyr
C19	Thr-Gly-Ser-Leu-Asn-Trp
C20	(Val, Pro, Val, Ser, Val, Glu, Gly, Tyr)Trp
C21	Asp-Scr-Ile-Thr-Hse

TABLE V

Amino Acid Composition of Pragment CB3

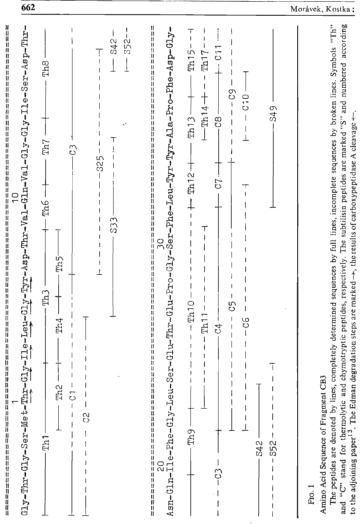
The values are given as number of amino acid residues in the molecule of the fragment. Arginine, lysine, histidine, a half-cystine were not found in the fragment.

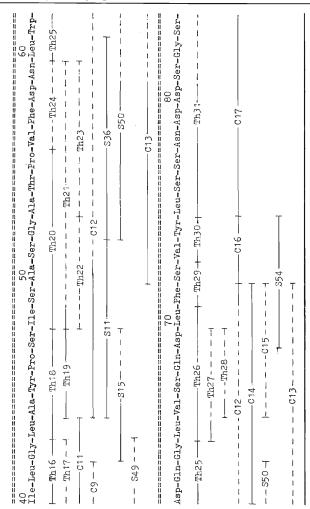
Fragment	Asp	Thr	Ser	Glu	Pro	Gly	Ala	Val	Ile	Leu	Tyr	Phe	Hse	Trp
CB3 (rcf. <sup>5</sup> ) In sequence of CB3														+ <sup>c</sup> 3

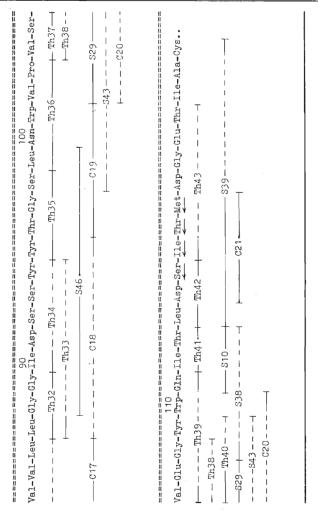
<sup>d</sup> Values extrapolated to zero time of hydrolysis; <sup>b</sup> values determined after 70-h hydrolysis; <sup>c</sup> the tryptophan content was determined qualitatively only. incompletely (fragment CB2) in which the N-terminal fragment CB4 is linked to fragment CB3 following in the chain. Fragment CB3 was obtained during the basic fractionation of the cyanogen bromide digest by gel filtration; this preparation containing contaminants was characterized<sup>1</sup> by the N-terminal sequence Thr-Gly-Ile--Leu-Gly-Tyr-... and by the C-terminal sequence Ser(lle,Thr)Hse. The carboxyl terminus of fragment CB3 is identical with the C-terminus of fragment CB2, determined as Ile-Thr-Hse. The isolation of the corresponding homoserine peptide from the chymotryptic digest of fragment CB2 (ref.<sup>10</sup>) permitted the carboxyl terminal region of fragment CB3 and CB2 to be formulated as Asp-Ser-Ile-Thr-Hse (ref.<sup>1</sup>). Fragment CB3 was purified by ion exchange chromatography<sup>5</sup>; the amino acid composition of this preparation is given in Table V. The tryptophan content (Table V) was not determined directly, it is known, however, from the analysis of fragment RAEP-tA 22 (2.9 residues, ref.<sup>2</sup>), whose terminal parts exceeding fragment CB3 do not contain tryptophan<sup>5</sup>. The N-terminal sequence of the fragment, Thr-Gly-Ile--Leu-Gly-Tyr-..., was verified by manual Edman degradation of pure fragment CB3. Because of the absence of lysine, arginine. and half-cystine (convertible into the aminoethylcysteine residue, sensitive to trypsin) in the part of the pepsin chain overlapping fragment CB3 this part of the chain was not cleaved by tryptic digestion of aminoethylated pepsin and a large fragment RAEP-tA 22 (ref.<sup>2</sup>) derived from this site was isolated. For the same reason tryptic hydrolysis could not be employed for sequential investigation of fragment CB3.

The amino acid sequence of the fragment (Fig. 1) was derived mainly from information afforded by chymotryptic, thermolytic, and subtilisin peptides. Data on the chymotryptic peptides obtained by the digestion of fragment CB3 were complemented by the alignment of peptides from the chymotryptic digest<sup>10</sup> of fragment CB2 (after elimination of peptides incorporated into the sequence<sup>5</sup> of fragment CB4) and, knowing the structures of the remaining cyanogen bromide fragments<sup>3-5</sup>. also by selected peptides from the chymotryptic hydrolysate of S-sulfo-pepsin<sup>11</sup>. Likewise, after the determination of the sequence of fragment CB4, the peptides remaining from the thermolytic digest<sup>10</sup> of fragment CB2 could be unambiguously incorporated into the region of CB3 and it became possible to also align selected peptides from the thermolysin digest of S-sulfo-pepsin<sup>12</sup>. Tryptic fragment RAEP-tA 22 (ref.<sup>2</sup>) served as a source for the preparation of subtilisin peptides. The subtilisin digest is described in more detail in the adjoining paper<sup>13</sup>; only selected peptides from this digest, complementing data obtained with other digests, are shown in the scheme in Fig. 1. In a number of cases the sequential characterization of the peptides (Table II and IV) was omitted wherever the information had been obtained with other peptides.

Unambiguous sequential overlaps (Fig. 1) permitted the amino acid sequence of the fragment to be arranged into four sections:







I	Thr-Gly-Ile	Ser-Val-Tyr	(res.	1-74)
П	Leu-Ser-Ser	Val-Val-Leu	(res.	75 - 86)
ш	Leu-Gly-Gly	Glp-Ile-Thr	(res.	87 - 113)
IV	Leu-Asp-Ser-Ile-Thr-Hse		(res.	114 - 119

We were not able to join together the above regions by overlapping peptides. The total sum of amino acid residues in these noneverlapping sequential regions equals the amino acid composition of fragment CB3 (Table V). The peptides from the individual enzymatic digests comply with this arrangement of sequential regions and do not suggest the existence of overlaps or, by contrast, a loss of peptides where these regions are linked to one another (residues 74/75, 86/87, and 113/114). The N-terminal sequence of region I is identical to the N-terminal sequence of fragment CB3; sequence IV, containing the homoserine residue, represents the carboxyl terminus of the fragment. The order of the middle regions II and III was established from the results of carboxypeptidase A degradation of fragments CB3 and CB2 (ref.<sup>1</sup>). During prolonged cleavage the quantity of threonine and isoleucine liberated exceeds the quantity of C-terminal homoserine; it may thus be assumed that the short sequential region IV (Leu-Asp-Ser-Ile-Thr-Hse) is preceded by a threonine and isoleucine residue. This assumption is in accordance with the carboxyl terminus of region III, ...Gln-Ile-Thr, since region II is C-terminated by ...Val-Val-Leu. As evidence supporting the location of region II after the N-terminal sequence I may be adducted the fact that the bond at the amino terminus of leucine No 75 is susceptible to cleavage by both chymotrypsin and bromosuccinimide<sup>18</sup>. This observation requires the presence of a tyrosine residue at the amino side of leucine 155 since the complete knowledge of the sequence around the three tryptophans of fragment CB3 excludes this residue as an alternative. Likewise, the sensitivity of the bond between residues 86 and 87 to chymotrypsin is compatible with the proposed bond Leu(86)-Leu(87).

The sequence of fragment CB2A, determined by Sepulveda and coworkers<sup>19</sup> is in complete agreement with our sequence of fragment CB3 (Fig. 1) which has already been reported in the paper describing the complete amino acid sequence of hog pepsin<sup>6</sup>; partial data pertaining to this part of the molecule were also published by Ostoslavskaya and coworkers<sup>20</sup>. Likewise the results reported by Revina and coworkers<sup>21</sup>, accounting for residues 100–119 of this fragment, confirm our sequence.

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